

# A rational strategy based on experimental designs to optimize parameters of a liquid chromatography-mass spectrometry analysis of complex matrices

Carole Reymond, Agnès Le Masle, Nadège Charon, Cyril Colas

# ▶ To cite this version:

Carole Reymond, Agnès Le Masle, Nadège Charon, Cyril Colas. A rational strategy based on experimental designs to optimize parameters of a liquid chromatography-mass spectrometry analysis of complex matrices. Talanta, 2019, 205, pp.120063. 10.1016/j.talanta.2019.06.063. hal-02196648

# HAL Id: hal-02196648 https://ifp.hal.science/hal-02196648

Submitted on 29 Jul 2019

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

- A rational strategy based on experimental designs to optimize parameters of a liquid chromatography-mass spectrometry analysis of complex matrices
- 3 Carole Reymond<sup>a</sup>, Agnès Le Masle<sup>a,\*</sup>, Cyril Colas<sup>b,c</sup>, Nadège Charon<sup>a</sup>
- 4 "IFP Energies nouvelles, Rond-point de l'échangeur de Solaize, BP 3, 69360 Solaize, France
- 5 <sup>b</sup> Institut de Chimie Organique et Analytique, Université d'Orléans, CNRS UMR 7311, Rue de Chartres, 45067
- 6 Orléans, France
- 7 Centre de Biophysique Moléculaire, CNRS UPR 4301, Université d'Orléans, rue Charles Sadron, 45071 Orléans,
- 8 France
- 9 \*Corresponding author. E-mail address: agnes.le-masle@ifpen.fr (A. Le Masle)

10 11

12

13

14 15

16 17

18

19

20

21

22

23

24 25

26

27

28

29

## Abstract

The ability of a method based on hyphenation of liquid chromatography and mass spectrometry to successfully separate, ionize and detect compounds in complex matrices is determined by a high number of factors controlled by the experimenter. Key steps to manage such hyphenation are focused on desolvation and ionization processes. In this study, a design of experiments approach was used to optimize decisive parameters (nebulizing, drying and sweep gas flow rates, ion transfer capillary voltage and temperature) for electrospray ionization and atmospheric pressure chemical ionization sources both in positive and negative modes. Central composite designs of 131 experiments were built in such a way as to cover rationally a sufficiently wide range of operating conditions and make possible the establishment of models. Each run was repeated three times to insure stable conditions of ionization and thus a satisfactory repeatability. Extracted ion chromatograms of twelve model compounds were integrated and entered as responses for experiment designs. Quadratic models for each standard allowed to take into account interactions between factors. Then responses were simultaneously maximized to achieve optimized factors. To illustrate the efficiency of this methodology, optimal conditions were applied to a lignocellulosic biomass fast pyrolysis oil. High sensitivity was obtained in LC/MS, especially for negative-ion mode electrospray which enabled identification of 5500 molecular formulae whereas direct introduction allowed to attribute twice less. In short, this study proposed a rational methodology to optimize ionization efficiency of a LC/MS analysis of complex mixtures.

- 30 Keywords:
- 31 LC-UV/MS; High resolution mass spectrometry; Design of experiments approach; Ionization efficiency;
- 32 Lignocellulosic biomass

## 34 1. Introduction

35 36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

61 62

63

64 65

66

67

68

69

70

71

72

73

74

75

76

77

Liquid chromatography (LC) hyphenated with mass spectrometry (MS) began to be studied much later than gas chromatography/mass spectrometry (GC/MS) because of the technical difficulties imposed by such hyphenation (desolvation in particular). It is only since thirty years that LC/MS has a real rise thanks to the development of ionization sources at atmospheric pressure. Despite the strong interest of the scientific community specially in metabolomics research, LC/MS is not systematically used as a routine technique [1]. Preliminary tests and an important step of optimization must be done when an analysis is set up. The main causes are the incompatibility of some chromatographic methods to mass spectrometry (solvent flow rate, additive, etc.) and the large number of parameters to manage in order to control the ionization efficiency inside the atmospheric pressure ionization source (API). Indeed, the signal obtained depends on several factors such as the flow rates of nebulising and drying gases, the sprayer position, the temperatures of the API and transfer capillary, capillary voltage, etc. Optimizing each of these parameters can be tedious and most of the time it is based on the experience that can have a regular user of a LC/MS system. Moreover, the procedure of varying one variable at a time does not guarantee the achievement of optimal conditions. Conversely, although barely used in the literature, the chemometric approach is based on a rational experimental model, which allows the simultaneous variation of all the experimental parameters. The use of an experimental plan also makes it possible to determine the number of necessary experiments and avoids the redundancies of information. It can also take into account the possible interactions between the different parameters. In our knowledge, only few publications used a design of experiments approach to optimize the ion source-mass spectrometry parameters mainly for pharmaceuticals issues [2-4]. Main of them focus on improving signal of target analytes for pharmaceuticals issues. Perrenoud et al. compared the sensitivity of a UHPSFC-ESI/MS/MS versus a UHPLC-ESI/MS/MS analysis for pharmaceutical compounds using a face-centered central composite design to optimize capillary voltage, desolvation temperature and drying gas flow rate [3].

In addition, LC/MS has become more and more interesting to analyze complex matrices containing for example heavy or thermosensitive products that cannot be analyzed by GC/MS. This is particularly true for oxygenated compounds coming from the conversion of lignocellulosic biomass [5–7]. In the present challenging energy transition context, biomass seems as a promising source to produce renewable energy, biofuels and chemical intermediates by mean of several transformation pathways [8,9]. As regards thermochemical pathway, lignocellulosic materials, mainly composed of lignin, cellulose and hemicellulose, are liquefied by fast pyrolysis at 400-450°C to produce bio-oils which might be further upgrated to biofuels [10,11]. The resulting liquid matrices are highly complex since they contain a wide diversity of oxygenated compounds (carboxylic acids, ketones, aldehydes, phenols, furans, carbohydrates, etc.). The large range of polarities and molecular weights but also heat sensitivity behavior of some compounds explain why chemical characterization of such liquids is still a challenge today [12]. Without prior derivatization step, GC and comprehensive twodimensional gas chromatography (GC×GC) could hardness detect compounds with a molecular weight higher than 200 g/mol [13–15]. However liquid chromatography (LC and LC×LC) appears to be a suitable technique thanks to its capacity to elute compounds with high polarity such as carbohydrates and/or high molecular weight [16,17]. More recently Fourier-transform ion cyclotron resonance mass spectrometry (FT-ICR/MS) has gained an increasing attention [18-23]. Highresolution mass spectrometry (HRMS) is a powerful technique for the analysis of many kinds of mixtures and provides an important list of chemical molecular formulae allowing a deeper understanding of the composition of bio-oils. Moreover, the use of several atmospheric pressure ionization sources is essential to detect a large range of molecules [24]. Some works have also demonstrated that additives like ammonium chloride or sodium hydroxide could enhance ionization efficiency of hemicellulosic and lignin derivative compounds [18,25,26]. However, one issue associated with complex mixture analysis based on mass spectrometry is signal suppression due to competition between analytes during the ionization process, especially with ESI. The hyphenation with liquid chromatography should theoretically reduce this phenomenon but also allow to differentiate isomers. Although a quite high number of papers using HRMS with sample direct introduction have been published on biomass samples [18,21,23–25], a much smaller number of studies was dedicated to the hyphenation of LC with HRMS [5,7,27].

In the present study, the aim was to develop an effective LC-UV/MS method to reach optimal signal. We have used a response surface methodology with central composite designs (CCD) in order to increase the ionization efficiency of model compounds related to bio-oils. LC/MS experiments were carried out using a LC/FT-ICR/MS hyphenated system. One experiment design were done for each ionization mode: electrospray ionization (ESI) and atmospheric pressure chemical ionization (APCI) both in positive and negative modes. Ahead the experiment designs, preliminary tests were made to fix some parameters like additives, split ratio and sprayer position). The objective was to develop a rational optimization of LC/MS method and thus to have a better control of the ionization efficiency. Our four methods were then applied to a lignocellulosic biomass pyrolysis oil.

# 2. Experimental

78

79

80

81

82

83

84

85

86

87

88

89

90

91

92

93

94

95

96

97

98

115

## 2.1. Chemicals

99 Methanol and formic acid were MS grade, purchased from VWR (Fontenay sous Bois, France). 100 Deionized water was produced by a Milli-Q water purifier (Millipore SAS, Molsheim, France). 1-(4-hydroxy-3-methoxyphenyl)ethan-1-one 101 Ammonium hydroxide, ammonium formate, 102 (acetovanillone), phenylethanone (acetophenone), 1-(4-ethylphenyl)ethanone ethylacetophenone), 1,3-cyclopentanedione, 2-phenylbenzo[h]chromen-4-one ( $\alpha$ -naphthoflavone), 103 104 2,5-hexanedione, 3-methyl-2-cyclohexen-1-one, 3-methyl-2-cyclopenten-1-one, 4-105 methylcyclohexanone, benzene-1-,2-diol (catechol), 2-methoxyphenol (guaiacol), 1,3-dimethoxy-2-106 hydroxybenzene (syringol), (3-phenoxyphenyl)methanol, 4-(hydroxymethyl)-2-methoxyphenol (vanillyl alcohol), butane-1,2,3,4-tetrol (erythritol), but-2-ene-1,4-dioic acid (fumaric acid), 107 108 octodecanoic acid (stearic acid), benzoic acid, 3-hydroxybenzoic acid, 4-hydroxybenzoic acid, 4-109 methylbenzoic acid (p-toluic acid), 4-hydroxy-3-methoxybenzoic acid (vanillic acid), 4-hydroxy-3,5-110 dimethoxybenzoic acid (syringic acid), benzaldehyde, 4-hydroxybenzaldehyde and 4-hydroxy-3-111 methoxybenzaldehyde (vanillin) were purchased from Sigma-Aldrich (Saint-Quentin-Fallavier, 112 France).

113 The fast pyrolysis bio-oil used in this study was produced from softwood sawdust and provided by IFP 114 Energies nouvelles (Solaize, France).

# 2.2. Sample preparation

116 Model mixtures used for CCD were prepared in methanol at a concentration of 1 mM for each compound. One mixture contained acetovanillone, acetophenone, 4-ethylacetophenone, 1,3-

cyclopentanedione, α-naphthoflavone, catechol, guaiacol, syringol, (3-phenoxyphenyl)methanol, vanillyl alcohol and erythritol. The second was composed of fumaric acid, stearic acid, benzoic acid, 3-hydroxybenzoic acid, 4-hydroxybenzoic acid, p-toluic acid, o-toluic acid, vanillic acid, syringic acid, benzaldehyde, 4-hydroxybenzaldehyde and vanillin (Table 1). The fast pyrolysis bio-oil was 10 times diluted in methanol before injection.

## 2.3. Instrumentation

All experiments were performed using an Agilent 1290 UHPLC system (Palo Alto, CA) consisting of a binary pump, an autosampler, a temperature-controlled column compartment and a photo diode array (PDA) detector (volume UV cell 1  $\mu$ L). The extra-column variance was measured at 9  $\mu$ L². The chromatographic system was hyphenated to a Thermo Scientific LTQ-FT-ICR (Bremen, Germany) composed of a linear ion trap and an ion cyclotron resonance cell in a 7 Tesla superconducting magnet. Ionization was carried out with the ESI and APCI sources both working in positive or negative modes.

# 2.4. High performance liquid chromatography/high-resolution mass spectrometry

For LC/MS analysis, 1  $\mu$ L of each sample was injected. Separation was achieved on a Kinetex C18 column (100 × 3 mm, particle size 2.6  $\mu$ m, Phenomenex, Le Pecq, France) at 30°C. The mobile phase solvents used were a 0.01% (v/v) formic acid in water (A) and a 0.01% (v/v) formic acid in methanol (B) at 600  $\mu$ L/min. A linear gradient was used as follows: 0.0 - 4.0 minutes, 1%B; 4.0 - 30.0 minutes, from 1% B to 99% B; 30.0 - 35.0 minutes, 99% B; 35.0 - 36.0 minutes, from 99% B to 1% B; 36.0-40.0 minutes, 1% B. UV signal was recorded from 210 to 400 nm.

To avoid excessive pressure on the UV cell and to be in optimal conditions for desolvation in ESI, LC flow was splited between UV detector and ESI source (Figure 1). Flow rates of nebulising, drying and sweep gases as well as transfer capillary temperature and voltage were optimized using a design of experiments approach (DoE). The spray voltage was -3.5 kV in negative mode and 4 kV in positive one. For APCI source, flow rates of nebulising and drying gases, as well as vaporizer temperature, transfer capillary temperature and voltage were optimized with a DoE. Discharge current was fixed at 20  $\mu$ A in positive and negative modes. No sweep gas was used. For both API sources, a delta of 25 V between tube lens voltage and transfer capillary voltage was kept constant. The sprayer position was also optimized.

In order to have at least 10 points for each chromatogram peak, a resolving power of 12500 at m/z 400 was used for the FT-ICR spectrometer. The mass range was set to m/z 92-1000 with 2 µscans averaging. External calibration was accomplished with CalMix+ and CalMix- (Thermo Scientific) in positive and negative mode respectively.

For Flow Injection Analysis (FIA), LC conditions were unchanged. MS conditions with APCI source were as follow: nebulising gas flow rate 40 AU, drying gas flow rate 20 AU, sweep gas flow rate 0 AU, capillary voltage -50V, capillary temperature 250°C, tube lens voltage -75V. MS conditions with ESI source were: nebulising gas flow rate 30 AU, drying gas flow rate 25 AU, sweep gas flow rate 0 AU, capillary voltage -30V, capillary temperature 350°C, tube lens voltage -55V.

## 2.5. Direct introduction (–)ESI/FT-ICR/MS

- 158 Sample was diluted in methanol (1:50; v/v) prior to the introduction by infusion (5  $\mu$ L/min). The
- number of μscans was set at 8 and 50 scans were accumulated with a resolution of 100000. Flow rate
- of nebulising gas was set to 5 AU. No drying and sweep gases were used. The spray voltage was -2.8
- 161 kV. Transfer capillary temperature, transfer capillary voltage and tube lens voltage were respectively
- 162 set to 275 °C, -35 V and -110 V.
- 163 2.6. Software and MS data processing
- 164 Extracted ion chromatograms (EICs) were traced and integrated with Xcalibur 2.1 software (Thermo
- 165 Scientific).
- 166 Construction of CCD, quadratic modellings of model molecules and optimization of MS parameters
- were done on Design Expert 10 software (Stat-Ease).
- 168 For molecular formulae calculation, homemade software was used. The following parameters were
- applied: elemental composition  $^{12}C_{1-100}$ ,  $^{1}H_{1-100}$ ,  $^{16}O_{1-100}$  and mass accuracy  $\leq 2$  ppm.

170

171

172

188

189

157

#### 3. Results and discussion

- 3.1. Methodology for optimizing desolvation and ionization parameters
- 173 The objective of this work was to develop a rational strategy to optimize parameters of a LC-UV/MS
- analysis of complex matrices coming from biomass such as bio-oils. When liquid chromatography is
- hyphenated to mass spectrometry, desolvation and ionization of molecules are critical steps. Indeed,
- when LC is used without MS, there is no restriction about additive/salt concentrations neither about
- 177 the LC flow rate which is directly related to chromatographic column's dimensions. To make LC/MS
- 178 hyphenation possible, many parameters have to be adjusted: LC flow rate, split ratio between LC and
- 179 MS system, nebulising, drying and sweep gas flow rates, transfer capillary temperature and voltage,
- tube lens voltage, vaporizer temperature for APCI, spray voltage or discharge current, API position
- are the main conditions to be investigated. To increase ionization efficiency, a make-up such as
- ammonium hydroxide in negative mode could also be added at the outlet of the LC system. In light of
- the large number of factors, to succeed this hyphenation and reach the real optimum, we propose a
- DoE methodology using central composite designs. Thereafter, a quadratic model was built, taking
- into account second order interactions.
- 186 This development was carried out on model compounds which are representative of bio-oils
- according to previous work [28]. Two mixtures were used (Table 1):
  - one for negative-ion ESI and APCI modes made of carboxylic acids and aldehydes
  - one for positive-ion ESI and APCI modes made of ketones and alcohols.
- 190 The main advantage to use model compounds was to cleverly optimize each ionization mode
- depending on the ability of compounds to accept or lose proton. Indeed, for example there is no
- 192 point to improve the ionization of carboxylic acids in positive mode while they are well ionized in
- 193 negative one thanks to the labile proton on the COOH function.

To build the four CCD, five significant parameters were chosen for each ionization mode in order to limit the number of experiments: flow rates of nebulising, drying and sweep gases, transfer capillary temperature and voltage (see part 3.2.). Tube lens voltage was associated with transfer capillary voltage by keeping a difference of 25 V between them in order to prevent ion source fragmentation. Other parameters which need manual adjustments were not included in the DoE and thus were optimized and fixed in advance: make-up and split of LC mobile phase at the entrance of mass spectrometer as well as the API source position. Impact of additives also called make-ups was also studied (see 3.1.). Chromatographic conditions were chosen to be compatible with mass spectrometry: for example concentration of formic acid was as low as possible but sufficient to keep analytes in their protonated form. Figure 1 illustrates the parameters which were either optimized in advance or with a DoE for ESI mode. To avoid any discharge and/or damage, authors decided to keep spray voltage at -3.5 kV and 4.0 kV for negative and positive ESI modes respectively. Moreover, according to previous publication about effects of several parameters linked to ion generation region on signal intensity using a similar ion source (electrospray ionization MAX2 source, Thermo Scientific); spray voltage was not found as a significant parameters at the opposite of the nebulising gas, capillary voltage and tube lens voltage. [29].

# 3.2. Adaptation of the LC downstream flow for MS compatibility

To optimize a LC/MS interface, it is primordial to well understand the process of ionization in order to focus on key parameters and adjust the flow rate and/or the composition of mobile phases to drive them compatible with ion source specificities.

In the case of ESI, the compounds diluted in the mobile phase are introduced by a capillary in an area at atmospheric pressure where an electric field of a few kV is applied. This electric field causes an accumulation of charges on the surface of the liquid causing the formation of a cone at the end of the capillary (Taylor cone). When coulombic repulsions exceed the surface tension of the liquid, charged droplets are then emitted from the tip of the cone. At this stage, several theories concerning the ionization process compete. The best known one is the residual charge theory introduced in 1968 by Dole *et al.* [30]. According to them, a succession of fissions of the droplets alternating with the evaporation of the solvent cause the release of ions in the gas phase. Conversely, according to other theories, like the model of ion evaporation (Iribarne and Thomson), the ions would be ejected from the droplets or even ejected directly from the Taylor cone [31]. In fact, the models may be complementary, and ionic evaporation may occur in the later stages of the residual charge theory [32,33]. It should be noted that the formation of ions in the gas phase often ends not in the atmospheric pressure area but at the interface of the latter and the next area at reduced pressure.

Unlike ESI for which ions are pre-formed in solution, APCI requires that the molecules go into the gas phase prior to ionization. For this reason, the ionization source is heated between 300 and 500°C (vaporizer temperature) depending on the flow rate and the composition of the liquid. The formed vapor then passes through a stream of electrons emitted by a corona discharge which creates plasma around the needle.

## 3.2.1. Flow splitting

LC flow rate was 600  $\mu$ L/min, which was too high for ESI. According to manufacturer, ESI could manage a flow of 1 mL/min, but optimal operation is obtained between 100 and 400  $\mu$ L/min. To reduce the flow rate, a T-split was installed after the chromatographic column to prevent UV cell

damages (Figure 1). Split ratio was calculated by divided flow rate at the outlet of the UV detector by the LC flow rate. The main drawback to split the mobile phase is solute dispersion, especially when a low split ratio has to be considered. Regarding theoretical considerations, total solute dispersion  $(\sigma^2_{total})$  can be assessed by peak variance coming from dispersion inside the column  $(\sigma^2_{col})$  and extracolumn dispersion  $(\sigma^2_{ext})$  (Eq 1). Extra-column dispersion results from injection, tubing, detector and split ratio (Eq 2) [34].

242 (1) 
$$\sigma_{total}^2 = \sigma_{col}^2 + \sigma_{ext}^2$$
  
243 (2)  $\sigma_{ext}^2 = \sigma_{injection}^2 + \sigma_{tubing}^2 + \sigma_{detector}^2 + \sigma_{split}^2$ 

To limit the loss of theoretical plates ( $N_{col}$ ), extra-column dispersion must be as low as possible. The ratio between  $\sigma^2_{col}$  and  $\sigma^2_{ext}$  ( $\beta^2$ ) represents the percentage of remaining plates. Here, we were interested in the ratio between  $\sigma^2_{col}$  and  $\sigma^2_{split}$ . For a compound with a retention factor at elution ( $k_e$ ) of 3,  $\sigma^2_{col}$  is equal to 187  $\mu L^2$  according to Eq 3.

248 (3) 
$$\sigma_{col}^2 = \frac{V_O^2 (1+k_e)^2}{N_{col}}$$

- 249 With  $V_0$  the column dead volume and  $N_{col}$  estimated by dividing the column length ( $L_{col}$ ) by 3 times the pore size ( $d_p$ ).
- To calculate the total dispersion, Dorsey-Foley equation was used (Eq.4).

252 (4) 
$$\sigma_{total}^2 = \frac{(As+1.25) F^2 W_{0.1}^2}{41.7}$$

255

256257

258

259

260

261262

263

264

265

270

- With F the mobile phase flow rate, As the peak asymmetry and  $W_{0.1}$  the peak width both at 10% of the peak height respectively.
  - By removing the chromatographic column,  $\sigma^2_{total}$  corresponds to  $\sigma^2_{ext}$ . In order to measure only the dispersion due to flow splitter, T-split was first removed to calculate the dispersion from injection, tubing and detection. Then this measured dispersion was subtracted to  $\sigma^2_{total}$  to obtain only the split dispersion. Several split ratios were tested by changing the LC flow rate and keeping constant the flow coming into the mass spectrometer. By doing this, we could consider that dispersion of mass spectrometer was constant. The variation of tubing's dispersion related to LC flow modification was considered negligible before split dispersion (from 100 to 600  $\mu$ L/min). Figure 2 shows the percentage of remaining plates according to split ratios. With a ratio of 0.20 (1:5), around 60% of plates were lost only because of the T-split which may lead to mix resolved peaks. To keep at least 70% of theoretical plates, a split ratio of minimum 0.45 (1:2.2) was needed which corresponds to a flow of 270  $\mu$ L/min directed into the mass spectrometer. Thus, this ratio was used later on.
- For APCI, no flow splitter was needed. Thanks to the tubular oven, APCI can handle higher flow rate than ESI. Moreover, APCI is a mass dependent ionization technique, which means a T-split would decrease the MS signal. At the opposite, ESI is concentration dependent, thus with adapted desolvation conditions, a flow splitter does not impact the MS signal [35].

## 3.2.2.Sprayer position

To optimize the source position of APCI and ESI, 4 model molecules were injected in FIA (Flow Injection Analysis) at a concentration of 1 mM. On the API used, four positions were possible from A

which is the closest position to the transfer ion capillary, to D the furthest one. Each EIC was integrated and measured areas as a function of the source position are plotted in Figure 3.

For APCI, the source position did not have a real impact on the areas of EIC and thus on the ionization efficiency of model compounds. For ESI, areas of compounds could be 5 times lower for position A compared to position D. So for all other experiments, API was fixed in D position. These observations were consistent with the ionization process of each API source. Indeed, in APCI source, the whole mobile phase is vaporized and the ionization occurs in the gas phase by the use of a corona discharge. At the opposite in ESI source, ionization takes place in liquid phase with the formation of a spray due to high voltage applied between the ESI needle and the ion sweep cone. Thus the position of ESI sprayer have a direct impact on the electric field (E) which becomes more intense by bringing the sprayer closer (Eq.5).

284 (5) 
$$E = \frac{V}{A \cdot r \cdot ln(\frac{4d}{r})}$$

With V the applied voltage, A an empirical constant estimated by Smith to be equal to 0.667, r the capillary emitter radius (cm) and d the distance between the ESI sprayer and the ground plane (cm) [36,37].

It is important to note that position source optimum depends on many other parameters such as nebulising gas flow rate or transfer ion capillary voltage. These parameters were optimized later with a DoE.

# 3.2.3. Influence of additives

It is well known that mobile phase composition plays an important role during desolvation and ionization steps [38–42]. In ESI, the solvent used must have a sufficient dielectric constant to form charged droplets as well as a low surface tension. Moreover, additives should be used with a concentration as low as possible. Indeed, ESI is a competitive ionization process. When several compounds are present in an electrospray droplet, they compete for a limited number of charged sites at the droplet surface, which can lead to ion suppression. In this study, the mobile phase contained 0.01% (v/v) of formic acid in order to keep carboxylic acids under their protonated forms during chromatographic separation. The measured pH of aqueous solvent was 3.2. For ESI positive-ion mode, formic acid was a good source of protons; however for negative-ion mode, a pH around 8 would be preferred to help anion formation in the liquid phase. To evaluate the effect on the mass signal of formic acid added in the mobile phase as well as two other additives, 4 model compounds (vanillic acid, syringic acid, vanillin and syringaldehyde) were injected in negative ESI using formic acid, ammonium hydroxide and ammonium formate at concentrations varying from 0.1  $\mu$ M to 50 mM.

Unexpectedly, formic acid did not deteriorate ionization efficiency at moderate concentrations (Figure 4). Loss of signal started around 1 mM. Between 0.1  $\mu$ M and 1 mM, ionization of carboxylic acids was not really impacted by formic acid, whereas aldehyde responses were exalted. This phenomenon is called "wrong-way-round ionization" [41,43]. ESI process is complex and many characteristics of mobile phase composition influence ionization process such as volatility, surface tension, conductivity, ionic strength, pH, gas-phase ion-molecule reactions, etc. Indeed, even if an analyte is ionized in liquid phase, ionization of neutral analytes present in the gas phase could also

occur by proton transfer reactions involving charged eluent species from solvents and/or additives. Such exaltation of aldehydes responses may be explained by the fact that ionization took place by gas-phase proton transfer or it may happen at the liquid-gas interface droplet. It is known that redox reactions occurring at ESI sprayer, *i.e.* reduction for negative-ion mode, create a significant difference of pH between the solution and the droplets [42]. Addition of protons from acids could facilitate chemical reduction and so decrease the pH at the surface of droplets [44]. Thus even if solutes were not ionized inside the mobile phase, modification of pH droplets could make ionization in liquid phase possible. However, this theory was not in agreement with the fact that carboxylic acids had an almost constant response although aldehydes shown an increasing of their responses, except if carboxylic acids were already well ionized without formic acid because of their low pKa.

Similarly to formic acid behavior, ammonium hydroxide and ammonium formate enhanced aldehyde responses, especially vanillin signal. These additives increased the pH, thus ionization could occur directly inside mobile phase. The main differences between ammonium hydroxide and formate were an enhancement less important for ammonium formate than ammonium hydroxide and an important ion suppression at high concentration of ammonium formate which did not occur for ammonium hydroxide. This last phenomenon could be due to the anionic part of ammonium formate HCOO<sup>-</sup> which is bigger than OH<sup>-</sup> for ammonium hydroxide and so had a better affinity for droplet surface leading to a competition with solutes at high concentrations.

During this study, 0.01% (v/v) of formic acid was added to mobile phase, which corresponds to 2.7 mM. These conditions were not unfavorable as predicted; thereby no other additive was used at the outlet of LC system.

# 3.3. Optimization of ESI and APCI desolvation conditions by mean of central composite design

CCD was chosen in this study in order to take into account second order interactions. For each ionization mode, a DoE was established including the 5 key parameters. For ESI, nebulising, drying and sweep gas flow rates, transfer ion capillary temperature and voltage were studied. For APCI, preliminary experiments showed that sweep gas flow rate did not influence ionization, thus it was excluded of the design and vaporizer temperature was added to the study. The investigated ranges for each variable are summarized in Table 2. These ranges had to be restricted to make the later second order modelling possible and in the same time sufficient to consider a relevant signal.

# Each CCD was composed of

- a factorial design with 2<sup>k</sup> experiments (k is the number of factors, here 5), each factorial point was repeated 3 times
- 2×k axial points also repeated 3 times
- 5 central points.

In this work, points were repeated whatever the conditions were, not only on central point as it usually done in literature. In this way repeatability of runs was estimated and unstable ionization conditions were pointed out, especially spray that may be not stable in the ESI source. In total, 131 injections of our model mixture were done for the four ionization modes. EIC were integrated for the 12 model compounds. Measured areas were used as responses of CCD and were exploited by creating one quadratic model per molecule. Models were constructed with 80% of experiments, the

- remaining 20% part was used as validating points. Last step was optimization of model compound
- 354 responses simultaneously with desirability function in order to obtain optimized conditions.
- Optimization of negative APCI is developed in this paper as example.
- 356 To correlate responses with factors, a second order model with interaction was fitted to the
- 357 experimental data. The general form of second order function is:
- 358 (6)  $y = \beta_0 + \sum_{i=1}^k \beta_i x_i + \sum_{i=1}^k \beta_{ii} x_i^2 + \sum_{i=1}^k \sum_{i>i}^k \beta_{ij} x_i x_i$
- Where y is the response,  $\beta_0$  the constant,  $\beta_i$  the first order coefficients,  $\beta_{ii}$  the quadratic coefficients
- and  $\beta_{ij}$  the interactive coefficients.
- 361 A model involving 5 factors requires calculation of 21 coefficients. First, to simplify the model, only
- 362 significant quadratic and interactive coefficients with a p-value lower than 0.1 were kept. Analysis of
- variance (ANOVA) for response surface reduced quadratic model of vanillic acid is shown in Table 3.
- The model F-value was equal to 58.42 indicating that the model is significant (P<0.05) and there was
- only 0.01% chance that the F-value this could occur due to noise. P-value less than 0.05 indicates that
- model terms are significant (with  $\alpha$  = 0.05). In this case A, B, D, E, AB, AC, AE, BC, BD, CD, CE, A<sup>2</sup>, C<sup>2</sup>
- 367 and D<sup>2</sup> were significant model terms. First order coefficient C, corresponding to capillary
- 368 temperature, was not significant; however quadratic coefficient C<sup>2</sup> and several interactive
- 369 coefficients with C were significant.
- 370 R-squared value was 0.9148 representing a good agreement between experimental results and the
- 371 model predictions. Moreover R-squared increased every time when an independent variable was
- added to the model, thus regression model containing more variables than another one is expected
- 373 to provide a better fit (main risk being model overfitting in this case). On the contrary, adjusted R-
- 374 squared increased only if a new term improved the model fit otherwise it decreased. Predicted R-
- 375 squared was used to determine how well model makes predictions. To calculate this value, a data
- point from the dataset was removed, then the regression equation was calculated, the last step was
- 377 to evaluate how well the model predicts the missing point. This procedure was done for all data
- points. In this case, predicted R-squared was 0.8786 and it was in good agreement with the adjusted
- 379 R-squared of 0.8992.
- 380 Once the model was built, several model evaluation plots could be traced. Figure 5 shows the
- 381 experimental data versus predicted value plot for the vanillic acid model. It can be observed that
- points are around the line, which means model fit to experimental data. Moreover, validation points,
- 383 notched points on the graph, were quite well predicted.
- 384 Interactions between factors could be represented with response surface. As an example, Figure 6
- 385 illustrates the response surface of ion transfer capillary temperature and ion transfer capillary
- voltage for vanillic acid. For low capillary voltage, the optimum capillary temperature was higher than
- 387 350°C while for high capillary voltage, a capillary temperature lower than 350°C was preferred.
- 388 When all the models for compounds were built, a desirability function was used to maximize at the
- 389 same time all the responses for predicted optimal conditions (Table 4). Figure 7 illustrates
- 390 perturbations at optimized conditions for the desirability and for two model compounds (vanillic acid
- and 4-hydroxybenzaldehyde). Optimal conditions will be applied for complex and unknown samples
- 392 containing similar analytes than chosen model compounds in this study. Thus a compromise was

made in order to obtain a sufficient signal for all model compounds without reaching the maximum. For example, the value of C factor, ion transfer capillary temperature, was optimized at 376°C, but it would be better for vanillic acid to use a lower temperature. As regards 4-hydroxybenzaldehyde, a slightly higher temperature would give a better response. Another remark concerns the stability of optimal conditions. The perturbation of desirability shows a plate, except for B factor (drying gas). This means that for example, if needed, smaller vaporizer temperature (E factor) could be used without having important consequence on signal. Globally, higher optimal nebulising and drying gas flow rates were obtained in positive modes than in negative ones. To facilitate desolvation, high temperatures were preferred in negative modes.

# 3.4. Application to a lignocellulosic biomass pyrolysis oil

Optimized ionization conditions were applied to a lignocellulosic biomass pyrolysis oil. LC-UV/HRMS analysis resulted from five complementary detections: UV, ESI and APCI in positive and negative modes. Moreover, all UV spectra were recorded from 210 to 400 nm. Molecular formulae were identified based only on carbon, hydrogen and oxygen elemental composition with an accuracy of 2 ppm (see part 2.5).

408 UV chromatogram at 254 nm and base peak chromatogram (BPC) of a lignocellulosic biomass 409 pyrolysis oil are shown in Figure 8 as examples. Optimized ESI negative-ion mode showed a high 410 sensitivity, especially between 10 and 30 minutes.

By integrating all BPC data coming from ESI negative-ion mode, two main zones are clearly distinguished on the mass spectrum (Figure 9). The first one includes m/z ratios varying between 100 and 450 whereas the second area ranges from 450 to 900 m/z ratios. When the pyrolytic oil was infused into the FT-ICR mass spectrometer without any chromatographic separation dimension, the second zone was not as well detected because of ion suppression.

When dealing with complex samples producing a large number of data to cope with, van Krevelen diagram is a good way to represent MS data. The highest number of molecular formulae was achieved with ESI negative-ion mode (ca. 5500 formulae). At the opposite only 500 formulae were identified with APCI negative-ion mode. Since they contain significant contents of sugars and other heat sensitive compounds, lignocellulosic bio-oils are known to be thermally unstable. So during MS analysis, compounds burned and formed a black substrate deposited on the tube oven of the APCI source because of the too high temperature (450°C). Experiments at 350°C did not improved the signal. ESI is a softer ionization source than APCI, thus it is more suitable for this kind of sample. However, APCI do not formed adducts during ionization process which can simplify data interpretation. In comparison to negative ionization mode, only 900 molecular formulae could be calculated with positive ionization mode (APCI and ESI). A van Krevelen diagram including all ionization modes was built (Figure 10). Compounds eluted with less than 20% of MeOH were represented in red, in blue those eluted between 20 and 80% of MeOH and in green with more than 80%. Red points correspond mainly to sugaric compounds coming from the degradation of cellulose and hemicellulose. These carbohydrates are very polar compounds and thus were non-retained on apolar stationary phase. Compounds showing a higher affinity with the stationary phase (blue points) correspond to pyrolytic lignin (phenolic compounds). Finally, green points were eluted with more than 80% of MeOH, they might correspond to lipids (fatty and resin acids).

It has to be noted that only 2500 molecular formulae could be attributed by direct introduction in negative ESI against 5500 with LC/MS due to matrix effects leading to ion suppression. Figure 9 already proved that heavy compounds (m/z > 500) were not well ionized with direct introduction. To target molecule class which suffer from ion suppression in infusion, the heteroatom class distribution for LC/MS and direct introduction, with oxygen families ranging from O<sub>0</sub> to O<sub>25</sub> is presented in Figure 11. It appears that molecules having low number of oxygen atoms (< 4) were not detected with direct introduction. Thus prior separation with liquid chromatography greatly enhances the detection of heavy and moderately polar molecules.

In addition to limit ion suppression, liquid chromatography hyphenated to mass spectrometry allowed to distinguish isomers which is not possible with direct introduction. An example is presented in Figure 12. EIC of m/z 259.0975  $\pm$  0.0005 in negative ESI corresponding to  $C_{15}H_{16}O_4$  shows four intense peaks and other smaller ones at different retention times indicating the presence of several isomers.

## 4. Conclusion

Despite LC-UV/HRMS analysis has shown a great interest for analysis of complex matrices since several years, no rational optimization methodology was proposed in order to obtain a sensitive but also controlled analytical method. In this work, a design of experiment was used to improve the desolvation and ionization steps of a system involving a reversed phase column hyphenated to a FT-ICR high resolution mass spectrometer. The effects of five experimental key parameters were studied using ESI and APCI sources in negative and positive-ion modes. In our study, more than 130 experiments were performed for each ionization mode, using a synthetic solution containing 12 oxygenated model compounds. This chemometric approach allowed to take into account all possible second order interactions, which will not be possible with the classic optimization 'one variable at a time'. Quadratic models were built for each model compound, then their responses were maximized at the same time to conclude about optimal conditions.

Moreover, some parameters (split ratio, sprayer position and additives) were investigated prior to carry out the design of experiments. As regards ESI analyses, a T-split was added in order to decrease the mobile phase flow rate at the entrance of the mass spectrometer. However dispersion induced by such system must be considered to keep enough theoretical plates and narrow peaks and achieve a satisfactory chromatographic separation. Signal detected in ESI was highly dependent on sprayer position, contrary to APCI. In our conditions (LC mobile phase containing 0.01% of formic acid), no supplementary additive was needed at the outlet of the chromatographic column.

Optimized conditions were applied to a lignocellulosic biomass fast pyrolysis oil to illustrate the efficiency of such a methodology. LC-FT-ICR/MS analysis in ESI negative-ion mode provides a detailed characterization of our complex bio-oil, delivering more than 5500 molecular formulae. Hyphenation coupling a chromatographic separation to high-resolution mass spectrometry was demonstrated as on one hand, LC first dimension enables to alleviate ion matrix effects and differentiate isomers having the same molecular formulae but different retention times, and on the other hand, precise and reliable molecular formulae are proposed for a large number of oxygenated compounds.

## Acknowledgments

The authors thank Abdelaziz Faraj from IFP Energies nouvelles for his help with design of experiments.

## 476 **5. References**

- 477 [1] S. Forcisi, F. Moritz, B. Kanawati, D. Tziotis, R. Lehmann, P. Schmitt-Kopplin, Liquid 478 chromatography-mass spectrometry in metabolomics research: mass analyzers in ultra high 479 pressure liquid chromatography coupling, J. Chromatogr. A 1292 (2013) 51–65. 480 https://doi.org/10.1016/j.chroma.2013.04.017.
- Parameters in Non-Steroidal Anti-Inflammatory and Analgesic Pharmaceuticals Analysis by a Design of Experiments Approach, J. Am. Soc. Mass Spectrom. 27 (10) (2016) 1703–1714. https://doi.org/10.1007/s13361-016-1459-0.
- 485 [3] A. Grand-Guillaume Perrenoud, J.-L. Veuthey, D. Guillarme, Coupling state-of-the-art supercritical fluid chromatography and mass spectrometry: From hyphenation interface optimization to high-sensitivity analysis of pharmaceutical compounds, J. Chromatogr. A 1339 (2014) 174–184. https://doi.org/10.1016/j.chroma.2014.03.006.
- 489 [4] M.D. Peris-Díaz, M.A. Sentandreu, E. Sentandreu, Multiobjective optimization of liquid 490 chromatography-triple-quadrupole mass spectrometry analysis of underivatized human urinary 491 amino acids through chemometrics, Anal. Bioanal. Chem. 410 (18) (2018) 4275–4284. 492 https://doi.org/10.1007/s00216-018-1083-x.
- 493 [5] T.M. Jarrell, C.L. Marcum, H. Sheng, B.C. Owen, C.J. O'Lenick, H. Maraun, J.J. Bozell, H.I. 494 Kenttämaa, Characterization of organosolv switchgrass lignin by using high performance liquid 495 chromatography/high resolution tandem mass spectrometry using hydroxide-doped negative-496 mode electrospray ionization, Green Chem 16 (2014)2713-2727. ion (5) 497 https://doi.org/10.1039/C3GC42355G.
- 498 [6] E. Kiyota, P. Mazzafera, Sawaya, Alexandra C H F, Analysis of soluble lignin in sugarcane by 499 ultrahigh performance liquid chromatography-tandem mass spectrometry with a do-it-yourself 500 oligomer database, Anal. Chem. 84 (16) (2012) 7015–7020. https://doi.org/10.1021/ac301112y.
- 501 [7] B.C. Owen, L.J. Haupert, T.M. Jarrell, C.L. Marcum, T.H. Parsell, M.M. Abu-Omar, J.J. Bozell, S.K. Black, H.I. Kenttämaa, High-performance liquid chromatography/high-resolution multiple stage tandem mass spectrometry using negative-ion-mode hydroxide-doped electrospray ionization for the characterization of lignin degradation products, Anal. Chem. 84 (14) (2012) 6000–6007. https://doi.org/10.1021/ac300762y.
- 506 [8] Ching Diego, Upgrading of biomass: alternative ways for biomass treatment, 2014.
- 507 [9] P.F.H. Harmsen, W.J.J. Huijgen, L.M. Bermudez Lopez, R.R.C. Bakker, Literature review of 508 physical and chemical pretreatment processes for lignocellulosic biomass, 2010 (accessed 3 509 November 2017).
- 510 [10] A.V. Bridgwater, Review of fast pyrolysis of biomass and product upgrading, Biomass and 511 Bioenergy 38 (2012) 68–94. https://doi.org/10.1016/j.biombioe.2011.01.048.
- 512 [11] D. Meier, B. van de Beld, A.V. Bridgwater, D.C. Elliott, A. Oasmaa, F. Preto, State-of-the-art of 513 fast pyrolysis in IEA bioenergy member countries, Renewable and Sustainable Energy Reviews 514 20 (2013) 619–641. https://doi.org/10.1016/j.rser.2012.11.061.
- 515 [12] N. Charon, J. Ponthus, D. Espinat, F. Broust, G. Volle, J. Valette, D. Meier, Multi-technique 516 characterization of fast pyrolysis oils, Journal of Analytical and Applied Pyrolysis 116 (2015) 18– 517 26. https://doi.org/10.1016/j.jaap.2015.10.012.

- 518 [13] J.H. Marsman, J. Wildschut, F. Mahfud, H.J. Heeres, Identification of components in fast 519 pyrolysis oil and upgraded products by comprehensive two-dimensional gas chromatography 520 and flame ionisation detection, J. Chromatogr. A 1150 (1-2) (2007) 21–27. 521 https://doi.org/10.1016/j.chroma.2006.11.047.
- 522 [14] K. Sipila, E. Kuoppala, L. Fagernas, A. Oasmaa, CHARACTERIZATION OF BIOMASS-BASED FLASH 523 PYROLYSIS OILS, Biomass and Bioenergy 14 (2) (1998) 103–113.
- 524 [15] M. Staš, D. Kubička, J. Chudoba, M. Pospíšil, Overview of Analytical Methods Used for Chemical 525 Characterization of Pyrolysis Bio-oil, Energy Fuels 28 (1) (2013) 385–402. 526 https://doi.org/10.1021/ef402047y.
- 527 [16] A. Le Masle, D. Angot, C. Gouin, A. D'Attoma, J. Ponthus, A. Quignard, S. Heinisch, Development 528 of on-line comprehensive two-dimensional liquid chromatography method for the separation of 529 biomass compounds, J. Chromatogr. A 1340 (2014) 90–98. 530 https://doi.org/10.1016/j.chroma.2014.03.020.
- 531 [17] D. Tomasini, F. Cacciola, F. Rigano, D. Sciarrone, P. Donato, M. Beccaria, E.B. Caramão, P. Dugo, 532 L. Mondello, Complementary analytical liquid chromatography methods for the characterization 533 of aqueous phase from pyrolysis of lignocellulosic biomasses, Anal. Chem. 86 (22) (2014) 11255– 534 11262. https://doi.org/10.1021/ac5038957.
- 535 [18] J. Hertzog, V. Carré, Y. Le Brech, A. Dufour, F. Aubriet, Toward Controlled Ionization Conditions 536 for ESI-FT-ICR-MS Analysis of Bio-Oils from Lignocellulosic Material, Energy Fuels 30 (7) (2016) 537 5729–5739. https://doi.org/10.1021/acs.energyfuels.6b00655.
- [19] J. Hertzog, V. Carré, Y. Le Brech, C.L. Mackay, A. Dufour, O. Mašek, F. Aubriet, Combination of electrospray ionization, atmospheric pressure photoionization and laser desorption ionization Fourier transform ion cyclotronic resonance mass spectrometry for the investigation of complex mixtures - Application to the petroleomic analysis of bio-oils, Anal. Chim. Acta 969 (2017) 26– 34. https://doi.org/10.1016/j.aca.2017.03.022.
- [20] Y. Liu, Q. Shi, Y. Zhang, Y. He, K.H. Chung, S. Zhao, C. Xu, Characterization of Red Pine Pyrolysis Bio-oil by Gas Chromatography–Mass Spectrometry and Negative-Ion Electrospray Ionization Fourier Transform Ion Cyclotron Resonance Mass Spectrometry, Energy Fuels 26 (7) (2012) 4532–4539. https://doi.org/10.1021/ef300501t.
- J.M. Jarvis, A.M. McKenna, R.N. Hilten, K.C. Das, R.P. Rodgers, A.G. Marshall, Characterization of
   Pine Pellet and Peanut Hull Pyrolysis Bio-oils by Negative-Ion Electrospray Ionization Fourier
   Transform Ion Cyclotron Resonance Mass Spectrometry, Energy Fuels 26 (6) (2012) 3810–3815.
   https://doi.org/10.1021/ef300385f.
- [22] E.A. Smith, S. Park, A.T. Klein, Y.J. Lee, Bio-oil Analysis Using Negative Electrospray Ionization:
   Comparative Study of High-Resolution Mass Spectrometers and Phenolic versus Sugaric
   Components, Energy Fuels 26 (6) (2012) 3796–3802. https://doi.org/10.1021/ef3003558.
- P.V. Abdelnur, B.G. Vaz, J.D. Rocha, de Almeida, Marlon B. B., M.A.G. Teixeira, R.C.L. Pereira, Characterization of Bio-oils from Different Pyrolysis Process Steps and Biomass Using High-Resolution Mass Spectrometry, Energy Fuels 27 (11) (2013) 6646–6654. https://doi.org/10.1021/ef400788v.
- 558 [24] M. Staš, J. Chudoba, M. Auersvald, D. Kubička, S. Conrad, T. Schulzke, M. Pospíšil, Application of 559 orbitrap mass spectrometry for analysis of model bio-oil compounds and fast pyrolysis bio-oils 560 from different biomass sources, Journal of Analytical and Applied Pyrolysis 124 (2017) 230–238. 561 https://doi.org/10.1016/j.jaap.2017.02.002.

- 562 [25] K.S. Boes, R.H. Narron, Y. Chen, S. Park, N.R. Vinueza, Characterization of biofuel refinery 563 byproduct via selective electrospray ionization tandem mass spectrometry, Fuel 188 (2017) 564 190–196. https://doi.org/10.1016/j.fuel.2016.10.016.
- [26] K.S. Boes, R.H. Narron, S. Park, N.R. Vinueza, Mass Spectrometry Exposes Undocumented Lignin Carbohydrate Complexes in Biorefinery Pretreatment Stream, ACS Sustainable Chem. Eng. 6 (8)
   (2018) 10654–10659. https://doi.org/10.1021/acssuschemeng.8b01986.
- J. Prothmann, M. Sun, P. Spégel, M. Sandahl, C. Turner, Ultra-high-performance supercritical fluid chromatography with quadrupole-time-of-flight mass spectrometry (UHPSFC/QTOF-MS)
   for analysis of lignin-derived monomeric compounds in processed lignin samples, Anal. Bioanal.
   Chem. 409 (30) (2017) 7049–7061. https://doi.org/10.1007/s00216-017-0663-5.
- 572 [28] J. Crepier, A. Le Masle, N. Charon, F. Albrieux, P. Duchene, S. Heinisch, Ultra-high performance 573 supercritical fluid chromatography hyphenated to atmospheric pressure chemical ionization 574 high resolution mass spectrometry for the characterization of fast pyrolysis bio-oils, J. 575 Chromatogr. В Analyt. Technol. Biomed. Life Sci. 1086 (2018)38-46. https://doi.org/10.1016/j.jchromb.2018.04.005. 576
- 577 [29] N. Lemonakis, A.-L. Skaltsounis, A. Tsarbopoulos, E. Gikas, Optimization of parameters affecting 578 signal intensity in an LTQ-orbitrap in negative ion mode: A design of experiments approach, 579 Talanta 147 (2016) 402–409. https://doi.org/10.1016/j.talanta.2015.10.009.
- 580 [30] M. Dole, L.L. Mack, R.L. Hines, R.C. Mobley, L.D. Ferguson, M.B. Alice, Molecular Beams of 581 Macroions, The Journal of Chemical Physics 49 (5) (1968) 2240–2249. https://doi.org/10.1063/1.1670391.
- 583 [31] J.V. Iribarne, B.A. Thomson, On the evaporation of small ions from charged droplets, The Journal of Chemical Physics 64 (1976) 2287. https://doi.org/10.1063/1.432536.
- [32] M. Wilm, Principles of electrospray ionization, Mol. Cell. Proteomics 10 (7) (2011) M111.009407.
   https://doi.org/10.1074/mcp.M111.009407.
- 587 [33] P. Kebarle, M. Peschke, On the mechanisms by which the charged droplets produced by 588 electrospray lead to gas phase ions, Anal. Chim. Acta 406 (1) (2000) 11–35. 589 https://doi.org/10.1016/S0003-2670(99)00598-X.
- 590 [34] M. Bernardin, F. Bessueille-Barbier, A. Le Masle, C.-P. Lienemann, S. Heinisch, Suitable interface 591 for coupling liquid chromatography to inductively coupled plasma-mass spectrometry for the 592 analysis of organic matrices. 1 Theoretical and experimental considerations on solute 593 dispersion, J. Chromatogr. A 1565 (2018) 68–80. https://doi.org/10.1016/j.chroma.2018.06.024.

594

595

596

597

- [35] D. Wolrab, P. Frühauf, C. Gerner, Direct coupling of supercritical fluid chromatography with tandem mass spectrometry for the analysis of amino acids and related compounds: Comparing electrospray ionization and atmospheric pressure chemical ionization, Anal. Chim. Acta 981 (2017) 106–115. https://doi.org/10.1016/j.aca.2017.05.005.
- 598 [36] J.W. Thompson, J.W. Eschelbach, R.T. Wilburn, J.W. Jorgenson, Investigation of electrospray 599 ionization and electrostatic focusing devices using a three-dimensional electrospray current 600 profiler, Am. Soc. Mass Spectrom. J. 16 (3) (2005)601 https://doi.org/10.1016/j.jasms.2004.11.012.
- 602 [37] D.P.H. Smith, The Electrohydrodynamic Atomization of Liquids, IEEE Trans. on Ind. Applicat. IA-603 22 (3) (1986) 527–535. https://doi.org/10.1109/TIA.1986.4504754.
- 604 [38] R. Dams, T. Benijts, W. Günther, W. Lambert, A. de Leenheer, Influence of the eluent composition on the ionization efficiency for morphine of pneumatically assisted electrospray,

- atmospheric-pressure chemical ionization and sonic spray, Rapid Commun. Mass Spectrom. 16 (11) (2002) 1072–1077. https://doi.org/10.1002/rcm.683.
- [39] B.A. Huffman, M.L. Poltash, C.A. Hughey, Effect of polar protic and polar aprotic solvents on
   negative-ion electrospray ionization and chromatographic separation of small acidic molecules,
   Anal. Chem. 84 (22) (2012) 9942–9950. https://doi.org/10.1021/ac302397b.
- [40] R. Kostiainen, A.P. Bruins, Effect of Solvent on Dynamic Range and Sensitivity in Pneumatically-assisted Electrospray (Ion Spray) Mass Spectrometry, Rapid Commun. Mass Spectrom. 10 (11) (1996) 1393–1399. https://doi.org/10.1002/(SICI)1097-0231(199608)10:11<1393:AID-RCM654>3.0.CO;2-V.
- 615 [41] R. Kostiainen, T.J. Kauppila, Effect of eluent on the ionization process in liquid chromatography-616 mass spectrometry, J. Chromatogr. A 1216 (4) (2009) 685–699. 617 https://doi.org/10.1016/j.chroma.2008.08.095.
- [42] Paul Kebarle/Liang Tang, From ions in solution to ions in the gas phase the mechanism of electrospray mass spectrometry, Anal. Chem. 65 (22) (1993) 972–986.
- [43] Z. Wu, W. Gao, M.A. Phelps, Di Wu, D.D. Miller, J.T. Dalton, Favorable effects of weak acids on
   negative-ion electrospray ionization mass spectrometry, Anal. Chem. 76 (3) (2004) 839–847.
   https://doi.org/10.1021/ac0351670.
- [44] N.B. Cech, C.G. Enke, Practical implications of some recent studies in electrospray ionization fundamentals, Mass Spectrom. Rev. 20 (6) (2001) 362–387. https://doi.org/10.1002/mas.10008.

# 626 Figure captions

625

- 627 Color should be used for Figure 1, Figure 3, Figure 4, Figure 5, Figure 8, Figure 10 and Figure 11.
- 628 **Table 1**: Model mixtures for negative-ion (left) and positive-ion (right) modes
- Figure 1: Key parameters for ESI detection optimized with DoE (red boxed text) and before DoE (purple boxed text)
- 631 Figure 2: Percentage of remaining plates according to the split ratio
- 632 Figure 3: Areas of EIC for 4 model compounds according to the ESI and APCI position (negative-ion
- 633 mode). [LC and MS conditions : see Experimental part]
- 634 Figure 4: Effect of formic acid, ammonium hydroxide and ammonium formate on negative-ion ESI
- 635 mode responses of 4 model compounds. The vertical axis represents the percentage ratio of peak
- area in the presence of additive to peak area in the absence of additive
- 637 **Table 2**: Central composite designs for ESI and APCI both in positive and negative modes
- 638 **Table 3**: ANOVA for response surface reduced quadratic model of vanillic acid in negative APCI
- 639 Figure 5: Experimental data versus predicted value plot for the vanillic acid model
- 640 Figure 6: Response surface plot of ion transfer capillary temperature and ion transfer capillary
- oltage. Nebulising gas flow rate, drying gas flow rate and vaporizer temperature were fixed at
- respectively 30 AU, 20 AU and 350°C in this representation.
- Figure 7: Perturbation graphs at optimized conditions for the desirability (left), vanillic acid (middle)
- and 4-hydroxybenaldehyde (right)
- Table 4: Optimized conditions resulting from model predictions for ESI and APCI sources
- 646 Figure 8: UV chromatogram at 254 nm (red) and BPC of negative ESI (blue) of a lignocellulosic
- 647 biomass pyrolysis oil
- 648 Figure 9: ESI Negative-ion mode mass spectra of a lignocellulosic biomass pyrolysis oil obtained with
- prior LC separation (top) and direct introduction (bottom)

- 650 **Figure 10**: Van Krevelen diagram of a lignocellulosic biomass pyrolysis oil.
- 651 Figure 11: Comparison of heteroatom class distributions between LC-(-)ESI/FT-ICR/MS and (-)ESI/FT-
- 652 ICR/MS
- Figure 12: Extraction ion chromatogram of a lignocellulosic biomass pyrolysis oil for of m/z 259.0975
- 654 +/- 0.0005