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A rational strategy based on experimental designs to optimize parameters of a liquid chromatography-mass spectrometry analysis of complex matrices

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- 10

11 Abstract

12 The ability of a method based on hyphenation of liquid chromatography and mass spectrometry to 13 successfully separate, ionize and detect compounds in complex matrices is determined by a high number of factors controlled by the experimenter. Key steps to manage such hyphenation are 14 15 focused on desolvation and ionization processes. In this study, a design of experiments approach was used to optimize decisive parameters (nebulizing, drying and sweep gas flow rates, ion transfer 16 17 capillary voltage and temperature) for electrospray ionization and atmospheric pressure chemical 18 ionization sources both in positive and negative modes. Central composite designs of 131 19 experiments were built in such a way as to cover rationally a sufficiently wide range of operating 20 conditions and make possible the establishment of models. Each run was repeated three times to 21 insure stable conditions of ionization and thus a satisfactory repeatability. Extracted ion 22 chromatograms of twelve model compounds were integrated and entered as responses for 23 experiment designs. Quadratic models for each standard allowed to take into account interactions between factors. Then responses were simultaneously maximized to achieve optimized factors. To 24 25 illustrate the efficiency of this methodology, optimal conditions were applied to a lignocellulosic 26 biomass fast pyrolysis oil. High sensitivity was obtained in LC/MS, especially for negative-ion mode 27 electrospray which enabled identification of 5500 molecular formulae whereas direct introduction 28 allowed to attribute twice less. In short, this study proposed a rational methodology to optimize 29 ionization efficiency of a LC/MS analysis of complex mixtures.

30 Keywords:

LC-UV/MS; High resolution mass spectrometry; Design of experiments approach; Ionization efficiency;
 Lignocellulosic biomass

33

34 1. Introduction

Liquid chromatography (LC) hyphenated with mass spectrometry (MS) began to be studied much 35 36 later than gas chromatography/mass spectrometry (GC/MS) because of the technical difficulties 37 imposed by such hyphenation (desolvation in particular). It is only since thirty years that LC/MS has a 38 real rise thanks to the development of ionization sources at atmospheric pressure. Despite the strong 39 interest of the scientific community specially in metabolomics research, LC/MS is not systematically 40 used as a routine technique [1]. Preliminary tests and an important step of optimization must be 41 done when an analysis is set up. The main causes are the incompatibility of some chromatographic 42 methods to mass spectrometry (solvent flow rate, additive, etc.) and the large number of parameters 43 to manage in order to control the ionization efficiency inside the atmospheric pressure ionization 44 source (API). Indeed, the signal obtained depends on several factors such as the flow rates of 45 nebulising and drying gases, the sprayer position, the temperatures of the API and transfer capillary, 46 capillary voltage, etc. Optimizing each of these parameters can be tedious and most of the time it is 47 based on the experience that can have a regular user of a LC/MS system. Moreover, the procedure of 48 varying one variable at a time does not guarantee the achievement of optimal conditions. 49 Conversely, although barely used in the literature, the chemometric approach is based on a rational 50 experimental model, which allows the simultaneous variation of all the experimental parameters. 51 The use of an experimental plan also makes it possible to determine the number of necessary 52 experiments and avoids the redundancies of information. It can also take into account the possible 53 interactions between the different parameters. In our knowledge, only few publications used a 54 design of experiments approach to optimize the ion source-mass spectrometry parameters mainly 55 for pharmaceuticals issues [2–4]. Main of them focus on improving signal of target analytes for 56 pharmaceuticals issues. Perrenoud et al. compared the sensitivity of a UHPSFC-ESI/MS/MS versus a 57 UHPLC-ESI/MS/MS analysis for pharmaceutical compounds using a face-centered central composite 58 design to optimize capillary voltage, desolvation temperature and drying gas flow rate [3].

59 In addition, LC/MS has become more and more interesting to analyze complex matrices containing 60 for example heavy or thermosensitive products that cannot be analyzed by GC/MS. This is particularly true for oxygenated compounds coming from the conversion of lignocellulosic biomass 61 62 [5–7]. In the present challenging energy transition context, biomass seems as a promising source to 63 produce renewable energy, biofuels and chemical intermediates by mean of several transformation pathways [8,9]. As regards thermochemical pathway, lignocellulosic materials, mainly composed of 64 65 lignin, cellulose and hemicellulose, are liquefied by fast pyrolysis at 400-450°C to produce bio-oils 66 which might be further upgrated to biofuels [10,11]. The resulting liquid matrices are highly complex 67 since they contain a wide diversity of oxygenated compounds (carboxylic acids, ketones, aldehydes, 68 phenols, furans, carbohydrates, etc.). The large range of polarities and molecular weights but also 69 heat sensitivity behavior of some compounds explain why chemical characterization of such liquids is 70 still a challenge today [12]. Without prior derivatization step, GC and comprehensive two-71 dimensional gas chromatography (GC×GC) could hardness detect compounds with a molecular 72 weight higher than 200 g/mol [13–15]. However liquid chromatography (LC and LC×LC) appears to be 73 a suitable technique thanks to its capacity to elute compounds with high polarity such as 74 carbohydrates and/or high molecular weight [16,17]. More recently Fourier-transform ion cyclotron 75 resonance mass spectrometry (FT-ICR/MS) has gained an increasing attention [18-23]. High-76 resolution mass spectrometry (HRMS) is a powerful technique for the analysis of many kinds of 77 mixtures and provides an important list of chemical molecular formulae allowing a deeper 78 understanding of the composition of bio-oils. Moreover, the use of several atmospheric pressure 79 ionization sources is essential to detect a large range of molecules [24]. Some works have also 80 demonstrated that additives like ammonium chloride or sodium hydroxide could enhance ionization 81 efficiency of hemicellulosic and lignin derivative compounds [18,25,26]. However, one issue 82 associated with complex mixture analysis based on mass spectrometry is signal suppression due to 83 competition between analytes during the ionization process, especially with ESI. The hyphenation 84 with liquid chromatography should theoretically reduce this phenomenon but also allow to 85 differentiate isomers. Although a quite high number of papers using HRMS with sample direct 86 introduction have been published on biomass samples [18,21,23-25], a much smaller number of 87 studies was dedicated to the hyphenation of LC with HRMS [5,7,27].

88 In the present study, the aim was to develop an effective LC-UV/MS method to reach optimal signal. 89 We have used a response surface methodology with central composite designs (CCD) in order to 90 increase the ionization efficiency of model compounds related to bio-oils. LC/MS experiments were 91 carried out using a LC/FT-ICR/MS hyphenated system. One experiment design were done for each 92 ionization mode: electrospray ionization (ESI) and atmospheric pressure chemical ionization (APCI) 93 both in positive and negative modes. Ahead the experiment designs, preliminary tests were made to 94 fix some parameters like additives, split ratio and sprayer position). The objective was to develop a 95 rational optimization of LC/MS method and thus to have a better control of the ionization efficiency. 96 Our four methods were then applied to a lignocellulosic biomass pyrolysis oil.

97 2. Experimental

98 2.1. Chemicals

99 Methanol and formic acid were MS grade, purchased from VWR (Fontenay sous Bois, France). 100 Deionized water was produced by a Milli-Q water purifier (Millipore SAS, Molsheim, France). 1-(4-hydroxy-3-methoxyphenyl)ethan-1-one 101 Ammonium hydroxide, ammonium formate, 102 (acetovanillone), phenylethanone (acetophenone), 1-(4-ethylphenyl)ethanone (4ethylacetophenone), 1,3-cyclopentanedione, 2-phenylbenzo[h]chromen-4-one (α -naphthoflavone), 103 104 2,5-hexanedione, 3-methyl-2-cyclohexen-1-one, 3-methyl-2-cyclopenten-1-one, 4-105 methylcyclohexanone, benzene-1-,2-diol (catechol), 2-methoxyphenol (guaiacol), 1,3-dimethoxy-2-106 hydroxybenzene (syringol), (3-phenoxyphenyl)methanol, 4-(hydroxymethyl)-2-methoxyphenol (vanillyl alcohol), butane-1,2,3,4-tetrol (erythritol), but-2-ene-1,4-dioic acid (fumaric acid), 107 108 octodecanoic acid (stearic acid), benzoic acid, 3-hydroxybenzoic acid, 4-hydroxybenzoic acid, 4-109 methylbenzoic acid (p-toluic acid), 4-hydroxy-3-methoxybenzoic acid (vanillic acid), 4-hydroxy-3,5-110 dimethoxybenzoic acid (syringic acid), benzaldehyde, 4-hydroxybenzaldehyde and 4-hydroxy-3-111 methoxybenzaldehyde (vanillin) were purchased from Sigma-Aldrich (Saint-Quentin-Fallavier, 112 France).

The fast pyrolysis bio-oil used in this study was produced from softwood sawdust and provided by IFPEnergies nouvelles (Solaize, France).

115 *2.2. Sample preparation*

116 Model mixtures used for CCD were prepared in methanol at a concentration of 1 mM for each 117 compound. One mixture contained acetovanillone, acetophenone, 4-ethylacetophenone, 1,3cyclopentanedione, α-naphthoflavone, catechol, guaiacol, syringol, (3-phenoxyphenyl)methanol,
vanillyl alcohol and erythritol. The second was composed of fumaric acid, stearic acid, benzoic acid,
3-hydroxybenzoic acid, 4-hydroxybenzoic acid, p-toluic acid, o-toluic acid, vanillic acid, syringic acid,
benzaldehyde, 4-hydroxybenzaldehyde and vanillin (Table 1). The fast pyrolysis bio-oil was 10 times
diluted in methanol before injection.

123 2.3. Instrumentation

All experiments were performed using an Agilent 1290 UHPLC system (Palo Alto, CA) consisting of a binary pump, an autosampler, a temperature-controlled column compartment and a photo diode array (PDA) detector (volume UV cell 1 μ L). The extra-column variance was measured at 9 μ L². The chromatographic system was hyphenated to a Thermo Scientific LTQ-FT-ICR (Bremen, Germany) composed of a linear ion trap and an ion cyclotron resonance cell in a 7 Tesla superconducting magnet. Ionization was carried out with the ESI and APCI sources both working in positive or negative modes.

131 2.4. High performance liquid chromatography/high-resolution mass spectrometry

For LC/MS analysis, 1 μ L of each sample was injected. Separation was achieved on a Kinetex C18 column (100 × 3 mm, particle size 2.6 μ m, Phenomenex, Le Pecq, France) at 30°C. The mobile phase solvents used were a 0.01% (v/v) formic acid in water (A) and a 0.01% (v/v) formic acid in methanol (B) at 600 μ L/min. A linear gradient was used as follows: 0.0 - 4.0 minutes, 1%B; 4.0 - 30.0 minutes, from 1% B to 99% B; 30.0 - 35.0 minutes, 99% B; 35.0 - 36.0 minutes, from 99% B to 1% B; 36.0-40.0 minutes, 1% B. UV signal was recorded from 210 to 400 nm.

138 To avoid excessive pressure on the UV cell and to be in optimal conditions for desolvation in ESI, LC 139 flow was splited between UV detector and ESI source (Figure 1). Flow rates of nebulising, drying and 140 sweep gases as well as transfer capillary temperature and voltage were optimized using a design of 141 experiments approach (DoE). The spray voltage was -3.5 kV in negative mode and 4 kV in positive 142 one. For APCI source, flow rates of nebulising and drying gases, as well as vaporizer temperature, 143 transfer capillary temperature and voltage were optimized with a DoE. Discharge current was fixed at 144 $20 \ \mu\text{A}$ in positive and negative modes. No sweep gas was used. For both API sources, a delta of $25 \ V$ 145 between tube lens voltage and transfer capillary voltage was kept constant. The sprayer position was 146 also optimized.

147 In order to have at least 10 points for each chromatogram peak, a resolving power of 12500 at m/z148 400 was used for the FT-ICR spectrometer. The mass range was set to m/z 92-1000 with 2 µscans 149 averaging. External calibration was accomplished with CalMix+ and CalMix- (Thermo Scientific) in 150 positive and negative mode respectively.

For Flow Injection Analysis (FIA), LC conditions were unchanged. MS conditions with APCI source were as follow: nebulising gas flow rate 40 AU, drying gas flow rate 20 AU, sweep gas flow rate 0 AU, capillary voltage -50V, capillary temperature 250°C, tube lens voltage -75V. MS conditions with ESI source were : nebulising gas flow rate 30 AU, drying gas flow rate 25 AU, sweep gas flow rate 0 AU, capillary voltage -30V, capillary temperature 350°C, tube lens voltage -55V.

156

157 2.5. Direct introduction (–)ESI/FT-ICR/MS

Sample was diluted in methanol (1:50; v/v) prior to the introduction by infusion (5 μL/min). The
number of μscans was set at 8 and 50 scans were accumulated with a resolution of 100000. Flow rate
of nebulising gas was set to 5 AU. No drying and sweep gases were used. The spray voltage was -2.8
kV. Transfer capillary temperature, transfer capillary voltage and tube lens voltage were respectively
set to 275 °C, -35 V and -110 V.

163 2.6. Software and MS data processing

164 Extracted ion chromatograms (EICs) were traced and integrated with Xcalibur 2.1 software (Thermo165 Scientific).

- 166 Construction of CCD, quadratic modellings of model molecules and optimization of MS parameters167 were done on Design Expert 10 software (Stat-Ease).
- 168 For molecular formulae calculation, homemade software was used. The following parameters were
- applied: elemental composition ${}^{12}C_{1-100}$, ${}^{11}H_{1-100}$, ${}^{16}O_{1-100}$ and mass accuracy ≤ 2 ppm.
- 170

171 3. Results and discussion

172 3.1. Methodology for optimizing desolvation and ionization parameters

173 The objective of this work was to develop a rational strategy to optimize parameters of a LC-UV/MS 174 analysis of complex matrices coming from biomass such as bio-oils. When liquid chromatography is 175 hyphenated to mass spectrometry, desolvation and ionization of molecules are critical steps. Indeed, 176 when LC is used without MS, there is no restriction about additive/salt concentrations neither about 177 the LC flow rate which is directly related to chromatographic column's dimensions. To make LC/MS 178 hyphenation possible, many parameters have to be adjusted: LC flow rate, split ratio between LC and 179 MS system, nebulising, drying and sweep gas flow rates, transfer capillary temperature and voltage, 180 tube lens voltage, vaporizer temperature for APCI, spray voltage or discharge current, API position 181 are the main conditions to be investigated. To increase ionization efficiency, a make-up such as ammonium hydroxide in negative mode could also be added at the outlet of the LC system. In light of 182 183 the large number of factors, to succeed this hyphenation and reach the real optimum, we propose a 184 DoE methodology using central composite designs. Thereafter, a quadratic model was built, taking into account second order interactions. 185

186 This development was carried out on model compounds which are representative of bio-oils 187 according to previous work [28]. Two mixtures were used (Table 1):

- 188 one for negative-ion ESI and APCI modes made of carboxylic acids and aldehydes
- 189 one for positive-ion ESI and APCI modes made of ketones and alcohols.

190 The main advantage to use model compounds was to cleverly optimize each ionization mode 191 depending on the ability of compounds to accept or lose proton. Indeed, for example there is no 192 point to improve the ionization of carboxylic acids in positive mode while they are well ionized in 193 negative one thanks to the labile proton on the COOH function. 194 To build the four CCD, five significant parameters were chosen for each ionization mode in order to 195 limit the number of experiments: flow rates of nebulising, drying and sweep gases, transfer capillary 196 temperature and voltage (see part 3.2.). Tube lens voltage was associated with transfer capillary 197 voltage by keeping a difference of 25 V between them in order to prevent ion source fragmentation. 198 Other parameters which need manual adjustments were not included in the DoE and thus were 199 optimized and fixed in advance: make-up and split of LC mobile phase at the entrance of mass 200 spectrometer as well as the API source position. Impact of additives also called make-ups was also 201 studied (see 3.1.). Chromatographic conditions were chosen to be compatible with mass 202 spectrometry: for example concentration of formic acid was as low as possible but sufficient to keep 203 analytes in their protonated form. Figure 1 illustrates the parameters which were either optimized in 204 advance or with a DoE for ESI mode. To avoid any discharge and/or damage, authors decided to keep 205 spray voltage at -3.5 kV and 4.0 kV for negative and positive ESI modes respectively. Moreover, 206 according to previous publication about effects of several parameters linked to ion generation region 207 on signal intensity using a similar ion source (electrospray ionization MAX2 source, Thermo 208 Scientific); spray voltage was not found as a significant parameters at the opposite of the nebulising 209 gas, capillary voltage and tube lens voltage. [29].

210 3.2. Adaptation of the LC downstream flow for MS compatibility

To optimize a LC/MS interface, it is primordial to well understand the process of ionization in order to focus on key parameters and adjust the flow rate and/or the composition of mobile phases to drive them compatible with ion source specificities.

214 In the case of ESI, the compounds diluted in the mobile phase are introduced by a capillary in an area 215 at atmospheric pressure where an electric field of a few kV is applied. This electric field causes an 216 accumulation of charges on the surface of the liquid causing the formation of a cone at the end of 217 the capillary (Taylor cone). When coulombic repulsions exceed the surface tension of the liquid, 218 charged droplets are then emitted from the tip of the cone. At this stage, several theories concerning 219 the ionization process compete. The best known one is the residual charge theory introduced in 1968 220 by Dole et al. [30]. According to them, a succession of fissions of the droplets alternating with the 221 evaporation of the solvent cause the release of ions in the gas phase. Conversely, according to other 222 theories, like the model of ion evaporation (Iribarne and Thomson), the ions would be ejected from 223 the droplets or even ejected directly from the Taylor cone [31]. In fact, the models may be 224 complementary, and ionic evaporation may occur in the later stages of the residual charge theory 225 [32,33]. It should be noted that the formation of ions in the gas phase often ends not in the 226 atmospheric pressure area but at the interface of the latter and the next area at reduced pressure.

Unlike ESI for which ions are pre-formed in solution, APCI requires that the molecules go into the gas phase prior to ionization. For this reason, the ionization source is heated between 300 and 500°C (vaporizer temperature) depending on the flow rate and the composition of the liquid. The formed vapor then passes through a stream of electrons emitted by a corona discharge which creates plasma around the needle.

232 *3.2.1.Flow splitting*

233 LC flow rate was 600 μ L/min, which was too high for ESI. According to manufacturer, ESI could 234 manage a flow of 1 mL/min, but optimal operation is obtained between 100 and 400 μ L/min. To 235 reduce the flow rate, a T-split was installed after the chromatographic column to prevent UV cell damages (Figure 1). Split ratio was calculated by divided flow rate at the outlet of the UV detector by the LC flow rate. The main drawback to split the mobile phase is solute dispersion, especially when a low split ratio has to be considered. Regarding theoretical considerations, total solute dispersion (σ^2_{total}) can be assessed by peak variance coming from dispersion inside the column (σ^2_{col}) and extracolumn dispersion (σ^2_{ext}) (Eq 1). Extra-column dispersion results from injection, tubing, detector and split ratio (Eq 2) [34].

242 (1)
$$\sigma_{total}^2 = \sigma_{col}^2 + \sigma_{ext}^2$$

243 (2) $\sigma_{ext}^2 = \sigma_{injection}^2 + \sigma_{tubing}^2 + \sigma_{detector}^2 + \sigma_{split}^2$

To limit the loss of theoretical plates (N_{col}), extra-column dispersion must be as low as possible. The ratio between σ^2_{col} and σ^2_{ext} (β^2) represents the percentage of remaining plates. Here, we were interested in the ratio between σ^2_{col} and σ^2_{split} . For a compound with a retention factor at elution (k_e) of 3, σ^2_{col} is equal to 187 μ L² according to Eq 3.

248 (3)
$$\sigma_{col}^2 = \frac{V_O^2 (1+k_e)^2}{N_{col}}$$

249 With V_0 the column dead volume and N_{col} estimated by dividing the column length (L_{col}) by 3 times 250 the pore size (d_p).

251 To calculate the total dispersion, Dorsey-Foley equation was used (Eq.4).

252 (4)
$$\sigma_{total}^2 = \frac{(As+1.25) F^2 W_{0.1}^2}{41.7}$$

253 With F the mobile phase flow rate, As the peak asymmetry and $W_{0.1}$ the peak width both at 10% of 254 the peak height respectively.

By removing the chromatographic column, σ^2_{total} corresponds to σ^2_{ext} . In order to measure only the 255 dispersion due to flow splitter, T-split was first removed to calculate the dispersion from injection, 256 257 tubing and detection. Then this measured dispersion was subtracted to σ^2_{total} to obtain only the split 258 dispersion. Several split ratios were tested by changing the LC flow rate and keeping constant the 259 flow coming into the mass spectrometer. By doing this, we could consider that dispersion of mass 260 spectrometer was constant. The variation of tubing's dispersion related to LC flow modification was considered negligible before split dispersion (from 100 to 600 µL/min). Figure 2 shows the 261 262 percentage of remaining plates according to split ratios. With a ratio of 0.20 (1:5), around 60% of 263 plates were lost only because of the T-split which may lead to mix resolved peaks. To keep at least 264 70% of theoretical plates, a split ratio of minimum 0.45 (1:2.2) was needed which corresponds to a 265 flow of 270 μ L/min directed into the mass spectrometer. Thus, this ratio was used later on.

For APCI, no flow splitter was needed. Thanks to the tubular oven, APCI can handle higher flow rate than ESI. Moreover, APCI is a mass dependent ionization technique, which means a T-split would decrease the MS signal. At the opposite, ESI is concentration dependent, thus with adapted desolvation conditions, a flow splitter does not impact the MS signal [35].

270 *3.2.2.Sprayer position*

To optimize the source position of APCI and ESI, 4 model molecules were injected in FIA (Flow Injection Analysis) at a concentration of 1 mM. On the API used, four positions were possible from A which is the closest position to the transfer ion capillary, to D the furthest one. Each EIC was integrated and measured areas as a function of the source position are plotted in Figure 3.

275 For APCI, the source position did not have a real impact on the areas of EIC and thus on the ionization 276 efficiency of model compounds. For ESI, areas of compounds could be 5 times lower for position A 277 compared to position D. So for all other experiments, API was fixed in D position. These observations 278 were consistent with the ionization process of each API source. Indeed, in APCI source, the whole 279 mobile phase is vaporized and the ionization occurs in the gas phase by the use of a corona 280 discharge. At the opposite in ESI source, ionization takes place in liquid phase with the formation of a 281 spray due to high voltage applied between the ESI needle and the ion sweep cone. Thus the position 282 of ESI sprayer have a direct impact on the electric field (E) which becomes more intense by bringing 283 the sprayer closer (Eq.5).

284 (5)
$$E = \frac{V}{A \cdot r \cdot ln\left(\frac{4d}{r}\right)}$$

With V the applied voltage, A an empirical constant estimated by Smith to be equal to 0.667, r the capillary emitter radius (cm) and d the distance between the ESI sprayer and the ground plane (cm) [36,37].

It is important to note that position source optimum depends on many other parameters such as
 nebulising gas flow rate or transfer ion capillary voltage. These parameters were optimized later with
 a DoE.

291 *3.2.3.Influence of additives*

292 It is well known that mobile phase composition plays an important role during desolvation and 293 ionization steps [38-42]. In ESI, the solvent used must have a sufficient dielectric constant to form 294 charged droplets as well as a low surface tension. Moreover, additives should be used with a 295 concentration as low as possible. Indeed, ESI is a competitive ionization process. When several 296 compounds are present in an electrospray droplet, they compete for a limited number of charged 297 sites at the droplet surface, which can lead to ion suppression. In this study, the mobile phase 298 contained 0.01% (v/v) of formic acid in order to keep carboxylic acids under their protonated forms 299 during chromatographic separation. The measured pH of aqueous solvent was 3.2. For ESI positive-300 ion mode, formic acid was a good source of protons; however for negative-ion mode, a pH around 8 301 would be preferred to help anion formation in the liquid phase. To evaluate the effect on the mass 302 signal of formic acid added in the mobile phase as well as two other additives, 4 model compounds 303 (vanillic acid, syringic acid, vanillin and syringaldehyde) were injected in negative ESI using formic 304 acid, ammonium hydroxide and ammonium formate at concentrations varying from 0.1 μ M to 50 305 mM.

306 Unexpectedly, formic acid did not deteriorate ionization efficiency at moderate concentrations 307 (Figure 4). Loss of signal started around 1 mM. Between 0.1 μM and 1 mM, ionization of carboxylic 308 acids was not really impacted by formic acid, whereas aldehyde responses were exalted. This 309 phenomenon is called "wrong-way-round ionization" [41,43]. ESI process is complex and many 310 characteristics of mobile phase composition influence ionization process such as volatility, surface 311 tension, conductivity, ionic strength, pH, gas-phase ion-molecule reactions, etc. Indeed, even if an 312 analyte is ionized in liquid phase, ionization of neutral analytes present in the gas phase could also 313 occur by proton transfer reactions involving charged eluent species from solvents and/or additives. 314 Such exaltation of aldehydes responses may be explained by the fact that ionization took place by 315 gas-phase proton transfer or it may happen at the liquid-gas interface droplet. It is known that redox 316 reactions occurring at ESI sprayer, *i.e.* reduction for negative-ion mode, create a significant difference 317 of pH between the solution and the droplets [42]. Addition of protons from acids could facilitate 318 chemical reduction and so decrease the pH at the surface of droplets [44]. Thus even if solutes were 319 not ionized inside the mobile phase, modification of pH droplets could make ionization in liquid 320 phase possible. However, this theory was not in agreement with the fact that carboxylic acids had an 321 almost constant response although aldehydes shown an increasing of their responses, except if 322 carboxylic acids were already well ionized without formic acid because of their low pKa.

323 Similarly to formic acid behavior, ammonium hydroxide and ammonium formate enhanced aldehyde 324 responses, especially vanillin signal. These additives increased the pH, thus ionization could occur 325 directly inside mobile phase. The main differences between ammonium hydroxide and formate were 326 an enhancement less important for ammonium formate than ammonium hydroxide and an 327 important ion suppression at high concentration of ammonium formate which did not occur for 328 ammonium hydroxide. This last phenomenon could be due to the anionic part of ammonium formate 329 HCOO⁻ which is bigger than OH⁻ for ammonium hydroxide and so had a better affinity for droplet surface leading to a competition with solutes at high concentrations. 330

During this study, 0.01% (v/v) of formic acid was added to mobile phase, which corresponds to 2.7 mM. These conditions were not unfavorable as predicted; thereby no other additive was used at the outlet of LC system.

334 3.3. Optimization of ESI and APCI desolvation conditions by mean of central composite design

CCD was chosen in this study in order to take into account second order interactions. For each ionization mode, a DoE was established including the 5 key parameters. For ESI, nebulising, drying and sweep gas flow rates, transfer ion capillary temperature and voltage were studied. For APCI, preliminary experiments showed that sweep gas flow rate did not influence ionization, thus it was excluded of the design and vaporizer temperature was added to the study. The investigated ranges for each variable are summarized in Table 2. These ranges had to be restricted to make the later second order modelling possible and in the same time sufficient to consider a relevant signal.

- 342 Each CCD was composed of
- a factorial design with 2^k experiments (k is the number of factors, here 5), each factorial
 point was repeated 3 times
- 345 2×k axial points also repeated 3 times
- 346 5 central points.

In this work, points were repeated whatever the conditions were, not only on central point as it usually done in literature. In this way repeatability of runs was estimated and unstable ionization conditions were pointed out, especially spray that may be not stable in the ESI source. In total, 131 injections of our model mixture were done for the four ionization modes. EIC were integrated for the 12 model compounds. Measured areas were used as responses of CCD and were exploited by creating one quadratic model per molecule. Models were constructed with 80% of experiments, the

- remaining 20% part was used as validating points. Last step was optimization of model compound responses simultaneously with desirability function in order to obtain optimized conditions. Optimization of negative APCI is developed in this paper as example.
- To correlate responses with factors, a second order model with interaction was fitted to the experimental data. The general form of second order function is:

358 (6)
$$y = \beta_0 + \sum_{i=1}^k \beta_i x_i + \sum_{i=1}^k \beta_{ii} x_i^2 + \sum_{i=1}^k \sum_{j>i}^k \beta_{ij} x_i x_j$$

359 Where y is the response, β_0 the constant, β_i the first order coefficients, β_{ii} the quadratic coefficients 360 and β_{ii} the interactive coefficients.

A model involving 5 factors requires calculation of 21 coefficients. First, to simplify the model, only 361 362 significant quadratic and interactive coefficients with a p-value lower than 0.1 were kept. Analysis of 363 variance (ANOVA) for response surface reduced quadratic model of vanillic acid is shown in Table 3. 364 The model F-value was equal to 58.42 indicating that the model is significant (P<0.05) and there was only 0.01% chance that the F-value this could occur due to noise. P-value less than 0.05 indicates that 365 366 model terms are significant (with α = 0.05). In this case A, B, D, E, AB, AC, AE, BC, BD, CD, CE, A², C² and D² were significant model terms. First order coefficient C, corresponding to capillary 367 temperature, was not significant; however quadratic coefficient C² and several interactive 368 369 coefficients with C were significant.

- 370 R-squared value was 0.9148 representing a good agreement between experimental results and the 371 model predictions. Moreover R-squared increased every time when an independent variable was 372 added to the model, thus regression model containing more variables than another one is expected 373 to provide a better fit (main risk being model overfitting in this case). On the contrary, adjusted R-374 squared increased only if a new term improved the model fit otherwise it decreased. Predicted R-375 squared was used to determine how well model makes predictions. To calculate this value, a data 376 point from the dataset was removed, then the regression equation was calculated, the last step was 377 to evaluate how well the model predicts the missing point. This procedure was done for all data 378 points. In this case, predicted R-squared was 0.8786 and it was in good agreement with the adjusted 379 R-squared of 0.8992.
- Once the model was built, several model evaluation plots could be traced. Figure 5 shows the experimental data *versus* predicted value plot for the vanillic acid model. It can be observed that points are around the line, which means model fit to experimental data. Moreover, validation points, notched points on the graph, were quite well predicted.
- Interactions between factors could be represented with response surface. As an example, Figure 6 illustrates the response surface of ion transfer capillary temperature and ion transfer capillary voltage for vanillic acid. For low capillary voltage, the optimum capillary temperature was higher than 350°C while for high capillary voltage, a capillary temperature lower than 350°C was preferred.
- 388 When all the models for compounds were built, a desirability function was used to maximize at the 389 same time all the responses for predicted optimal conditions (Table 4). Figure 7 illustrates 390 perturbations at optimized conditions for the desirability and for two model compounds (vanillic acid 391 and 4-hydroxybenzaldehyde). Optimal conditions will be applied for complex and unknown samples 392 containing similar analytes than chosen model compounds in this study. Thus a compromise was

393 made in order to obtain a sufficient signal for all model compounds without reaching the maximum. 394 For example, the value of C factor, ion transfer capillary temperature, was optimized at 376°C, but it 395 would be better for vanillic acid to use a lower temperature. As regards 4-hydroxybenzaldehyde, a 396 slightly higher temperature would give a better response. Another remark concerns the stability of 397 optimal conditions. The perturbation of desirability shows a plate, except for B factor (drying gas). 398 This means that for example, if needed, smaller vaporizer temperature (E factor) could be used 399 without having important consequence on signal. Globally, higher optimal nebulising and drying gas 400 flow rates were obtained in positive modes than in negative ones. To facilitate desolvation, high 401 temperatures were preferred in negative modes.

402 3.4. Application to a lignocellulosic biomass pyrolysis oil

403 Optimized ionization conditions were applied to a lignocellulosic biomass pyrolysis oil. LC-UV/HRMS 404 analysis resulted from five complementary detections: UV, ESI and APCI in positive and negative 405 modes. Moreover, all UV spectra were recorded from 210 to 400 nm. Molecular formulae were 406 identified based only on carbon, hydrogen and oxygen elemental composition with an accuracy of 2 407 ppm (see part 2.5).

408 UV chromatogram at 254 nm and base peak chromatogram (BPC) of a lignocellulosic biomass 409 pyrolysis oil are shown in Figure 8 as examples. Optimized ESI negative-ion mode showed a high 410 sensitivity, especially between 10 and 30 minutes.

By integrating all BPC data coming from ESI negative-ion mode, two main zones are clearly distinguished on the mass spectrum (Figure 9). The first one includes *m/z* ratios varying between 100 and 450 whereas the second area ranges from 450 to 900 *m/z* ratios. When the pyrolytic oil was infused into the FT-ICR mass spectrometer without any chromatographic separation dimension, the second zone was not as well detected because of ion suppression.

416 When dealing with complex samples producing a large number of data to cope with, van Krevelen 417 diagram is a good way to represent MS data. The highest number of molecular formulae was 418 achieved with ESI negative-ion mode (ca. 5500 formulae). At the opposite only 500 formulae were 419 identified with APCI negative-ion mode. Since they contain significant contents of sugars and other 420 heat sensitive compounds, lignocellulosic bio-oils are known to be thermally unstable. So during MS 421 analysis, compounds burned and formed a black substrate deposited on the tube oven of the APCI 422 source because of the too high temperature (450°C). Experiments at 350°C did not improved the 423 signal. ESI is a softer ionization source than APCI, thus it is more suitable for this kind of sample. 424 However, APCI do not formed adducts during ionization process which can simplify data 425 interpretation. In comparison to negative ionization mode, only 900 molecular formulae could be 426 calculated with positive ionization mode (APCI and ESI). A van Krevelen diagram including all 427 ionization modes was built (Figure 10). Compounds eluted with less than 20% of MeOH were 428 represented in red, in blue those eluted between 20 and 80% of MeOH and in green with more than 429 80%. Red points correspond mainly to sugaric compounds coming from the degradation of cellulose 430 and hemicellulose. These carbohydrates are very polar compounds and thus were non-retained on 431 apolar stationary phase. Compounds showing a higher affinity with the stationary phase (blue points) 432 correspond to pyrolytic lignin (phenolic compounds). Finally, green points were eluted with more 433 than 80% of MeOH, they might correspond to lipids (fatty and resin acids).

434 It has to be noted that only 2500 molecular formulae could be attributed by direct introduction in 435 negative ESI against 5500 with LC/MS due to matrix effects leading to ion suppression. Figure 9 436 already proved that heavy compounds (m/z > 500) were not well ionized with direct introduction. To 437 target molecule class which suffer from ion suppression in infusion, the heteroatom class distribution 438 for LC/MS and direct introduction, with oxygen families ranging from O_0 to O_{25} is presented in Figure 439 11. It appears that molecules having low number of oxygen atoms (< 4) were not detected with 440 direct introduction. Thus prior separation with liquid chromatography greatly enhances the detection 441 of heavy and moderately polar molecules.

442 In addition to limit ion suppression, liquid chromatography hyphenated to mass spectrometry 443 allowed to distinguish isomers which is not possible with direct introduction. An example is 444 presented in Figure 12. EIC of m/z 259.0975 ± 0.0005 in negative ESI corresponding to $C_{15}H_{16}O_4$ 445 shows four intense peaks and other smaller ones at different retention times indicating the presence 446 of several isomers.

447 4. Conclusion

Despite LC-UV/HRMS analysis has shown a great interest for analysis of complex matrices since 448 449 several years, no rational optimization methodology was proposed in order to obtain a sensitive but 450 also controlled analytical method. In this work, a design of experiment was used to improve the 451 desolvation and ionization steps of a system involving a reversed phase column hyphenated to a FT-452 ICR high resolution mass spectrometer. The effects of five experimental key parameters were studied 453 using ESI and APCI sources in negative and positive-ion modes. In our study, more than 130 454 experiments were performed for each ionization mode, using a synthetic solution containing 12 455 oxygenated model compounds. This chemometric approach allowed to take into account all possible 456 second order interactions, which will not be possible with the classic optimization 'one variable at a 457 time'. Quadratic models were built for each model compound, then their responses were maximized 458 at the same time to conclude about optimal conditions.

Moreover, some parameters (split ratio, sprayer position and additives) were investigated prior to carry out the design of experiments. As regards ESI analyses, a T-split was added in order to decrease the mobile phase flow rate at the entrance of the mass spectrometer. However dispersion induced by such system must be considered to keep enough theoretical plates and narrow peaks and achieve a satisfactory chromatographic separation. Signal detected in ESI was highly dependent on sprayer position, contrary to APCI. In our conditions (LC mobile phase containing 0.01% of formic acid), no supplementary additive was needed at the outlet of the chromatographic column.

466 Optimized conditions were applied to a lignocellulosic biomass fast pyrolysis oil to illustrate the 467 efficiency of such a methodology. LC-FT-ICR/MS analysis in ESI negative-ion mode provides a detailed 468 characterization of our complex bio-oil, delivering more than 5500 molecular formulae. Hyphenation 469 coupling a chromatographic separation to high-resolution mass spectrometry was demonstrated as 470 on one hand, LC first dimension enables to alleviate ion matrix effects and differentiate isomers 471 having the same molecular formulae but different retention times, and on the other hand, precise 472 and reliable molecular formulae are proposed for a large number of oxygenated compounds.

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476 5. References

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- 625
- 626 Figure captions
- 627 Color should be used for Figure 1, Figure 3, Figure 4, Figure 5, Figure 8, Figure 10 and Figure 11.
- **Table 1**: Model mixtures for negative-ion (left) and positive-ion (right) modes
- 629 Figure 1: Key parameters for ESI detection optimized with DoE (red boxed text) and before DoE
- 630 (purple boxed text)
- 631 Figure 2 : Percentage of remaining plates according to the split ratio
- 632 Figure 3: Areas of EIC for 4 model compounds according to the ESI and APCI position (negative-ion
- 633 mode). [LC and MS conditions : see Experimental part]
- 634 Figure 4: Effect of formic acid, ammonium hydroxide and ammonium formate on negative-ion ESI
- mode responses of 4 model compounds. The vertical axis represents the percentage ratio of peak
- area in the presence of additive to peak area in the absence of additive
- 637 Table 2: Central composite designs for ESI and APCI both in positive and negative modes
- 638 Table 3: ANOVA for response surface reduced quadratic model of vanillic acid in negative APCI
- 639 Figure 5: Experimental data versus predicted value plot for the vanillic acid model
- 640 Figure 6: Response surface plot of ion transfer capillary temperature and ion transfer capillary
- voltage. Nebulising gas flow rate, drying gas flow rate and vaporizer temperature were fixed at
 respectively 30 AU, 20 AU and 350°C in this representation.
- 643 **Figure 7**: Perturbation graphs at optimized conditions for the desirability (left), vanillic acid (middle)
- 644 and 4-hydroxybenaldehyde (right)
- 645 **Table 4**: Optimized conditions resulting from model predictions for ESI and APCI sources
- Figure 8: UV chromatogram at 254 nm (red) and BPC of negative ESI (blue) of a lignocellulosicbiomass pyrolysis oil
- 648 **Figure 9**: ESI Negative-ion mode mass spectra of a lignocellulosic biomass pyrolysis oil obtained with
- 649 prior LC separation (top) and direct introduction (bottom)

- 650 **Figure 10**: Van Krevelen diagram of a lignocellulosic biomass pyrolysis oil.
- 651 Figure 11: Comparison of heteroatom class distributions between LC-(-)ESI/FT-ICR/MS and (-)ESI/FT-
- 652 ICR/MS
- **Figure 12**: Extraction ion chromatogram of a lignocellulosic biomass pyrolysis oil for of *m*/z 259.0975
- 654 +/- 0.0005